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Pharmaceutical composition for treating
papillomavirus tumors and infection

E1 >

The subject of the present invention is a pharmaceutical composition intended for the treatment or prevention of lesions associated with papillomaviruses and more particularly with human papillomaviruses (HPV) types 16, 18, 31, 33 and 45.

Papillomaviruses are DNA viruses possessing a circular genome of about 7900 base pairs surrounded by a protein capsid. A number of bovine (BPV) and human (HPV) papillomavirus types have been identified and their genome sequenced (Pfister, 1987, in *The papovaviridae : The Papillomaviruses* (Salzman and Howley publishing) Plenum Press, New York, p 1-38). It comprises an early region and a late region. The late region contains two reading frames L1 and L2 which code for the major components of the capsid. The early region contains at least the reading frames E1, E2, E4, E5, E6 and E7. The E1 and E2 expression products regulate viral replication and the expression of the viral genes whereas those of the E5, E6 and E7 regions are involved in the processes of oncogenic transformation of infected cells. Indeed, it has been shown experimentally that the BPV-1 E5 protein can transform cells in vitro (Schlegel et al., 1986, Science 233, 464-467). The BPV-1 E6 and HPV-16 E7 proteins are involved in the induction and maintenance of oncogenic transformation. The transforming power of E7 has been demonstrated for HPV-16 and HPV-18 (Kanda et al., 1988, J. Virol. 62, 610-613; Vousden et al., 1988, Oncogene Res. 3, 1-9; Bedell et al., 1987, J. Virol. 61, 3635-3640). No function has been demonstrated in E4.

In man, HPVs are associated with pathologies ranging from benign infection of the skin to warts and to malignant tumors. These viruses are highly specific for the epithelium of the epidermis of the genital, oral and respiratory tracts. The epidemiological data strongly suggest the role of certain strains in cancer of the neck

of the uterus and of the lower pathologies, in particular HPV-16 and -18 and to a lesser degree HPV-31, -33 and -45. Cancer of the neck of the uterus is the second cause of female cancer worldwide. According to the WHO, 460,000 new cases are recorded each year, of which at least 200,000 cases are clearly associated with HPV. A series of studies demonstrates the transforming role of these viruses, their specific integration into the genome of neoplastic cells, their gene activity in cancer cells and the importance of the expression of the E6 and E7 early genes in maintaining the malignant phenotype of HPV-positive neoplastic cells (Monsenego, J. Impact Medecin, March 11, 1994).

Pathological conditions associated with HPV viruses give rise to a therapeutic problem on account of their persistent and recurrent nature. Many approaches have already been used in the treatment of these diseases, such as surgery, chemotherapy, antiviral agents and immunotherapy.

In this regard, European Patent EP 0,462,187 describes a vaccination approach using papillomavirus early genes to establish immunity against tumors resulting from the integration of the HPV genome into the cellular DNA and in which the capsid proteins are no longer expressed. Application WO 93/02184 teaches a therapeutic approach based on the use of the capsid antigens as immunogenic agents. These documents do not suggest the possibility of combining the preventive effect offered by the early polypeptides and the curative effect conferred by the late polypeptides of papilloma-viruses to generate compositions suited to all the serious pathological conditions caused by HPVs. Moreover, during the past few years, it has been proposed to use polypeptides having immunostimulatory activity with the aim of activating the T cells with a result which is beneficial to a greater or lesser degree depending on the pathological conditions targeted (see for example WO 96/11279).

The present invention relates more precisely to

5 a preparation based on a mixture of antigens from the
early and late regions of a papillomavirus or a vector
expressing them simultaneously, with the aim of
establishing lasting immunity against infected cells. The
10 candidate vaccines provided within the framework of the
present invention can be used for preventive purposes
(immunoprophylaxy) to limit the development or propaga-
tion of viral infection to the neighboring tissues or for
curative purposes (immunotherapy) to prevent or reduce
15 tumor progression in infected patients. The use of the
capsid antigens will induce the production of antibodies
against the antigenic epitopes located at the surface of
the viral particles, preventing the infection from
becoming established for a lengthy period. The use of the
20 early proteins will make it possible to induce immunity
against the cells infected after integration of the viral
DNA.

The present invention also provides a preparation
combining a polypeptide from a papillomavirus and an
25 immunostimulatory molecule. One of the advantages of such
a composition is that it combines the specific immunity
induced by the viral antigens and the aspecific immunity
induced by the immunostimulatory molecule designed to
reinforce the specific response.

30 The aim of the present invention is to make
available to the public pharmaceutical compositions
allowing the treatment of HPV infections and more
particularly serious pathologies such as cancer of the
neck of the uterus, having improved efficacy compared
with the prior art compositions.

Accordingly, the subject of the present invention
is a pharmaceutical composition intended for the treat-
ment or prevention of a papillomavirus infection or
tumor, which comprises, as therapeutic agents:

- 35 (1) at least one polypeptide from the early region
of a papillomavirus and at least one poly-
peptide from the late region of a papilloma-
virus,
(2) at least one polypeptide from the early region

of a papillomavirus, at least one polypeptide from the late region of a papillomavirus and at least one polypeptide having immunostimulatory activity, or

- 5 (3) at least one polypeptide from an early or late region of a papillomavirus and at least one polypeptide having immunostimulatory activity.

In general, the term polypeptide refers to all or part of the native polypeptide, to a chimeric polypeptide
10 resulting from the fusion of sequences of different origins or to a variant characterized by at least one mutation (deletion, insertion and/or substitution) of an amino acid. More particularly, a polypeptide in use within the framework of the present invention has, in
15 particular, an amino acid sequence whose degree of similarity with the sequence of the native protein is greater than 75%, advantageously greater than 85% and, preferably, greater than 95%. The degree of similarity can be easily calculated with the aid of an appropriate
20 computer program or by aligning the sequences so as to obtain the maximum degree of homology and by counting the number of positions in which the amino acids of the two sequences are found to be identical in relation to the total number of positions. The sequence of the HPV-16 and
25 HPV-18 genomes is disclosed in Genebank with accession numbers K02718 and X05015 respectively.

As recalled earlier, the genome of viruses of the papillomavirus family, in particular BPV and HPV, codes for at least 8 polypeptides, two late polypeptides L1 and
30 L2 comprising the viral capsid and 6 early polypeptides (E1, E2, E4, E5, E6 and E7) involved in the regulation and maintenance of the viral genome and in the transformation of the infected cells.

Although all the early proteins of a papilloma-
35 virus can be used within the framework of the present invention, the choice is made advantageously to use a polypeptide derived from the E6 protein, from the E7 protein or from the E6 and E7 proteins. It may be advantageous to use a nononcogenic variant mutated at the

level of the regions involved in the process of transformation of the infected cells. Such variants are described in the literature (Munger et al., 1989, EMBO J. 8, 4099-4105; Crook et al., 1991, Cell 67, 547-556; Heck
5 et al., 1992, Proc. Natl. Acad. Sci. USA 89, 4442-4446; Phelps et al., 1992, J. Virol. 66, 2418-2427).

A preferred polypeptide from the late region of a papillomavirus is derived from the L1 protein, from the L2 protein or from the L1 and L2 proteins.

10 According to the particularly advantageous embodiments (2) and (3), a composition according to the invention also comprises a polypeptide having an immunostimulatory activity. "Immunostimulatory" is understood
15 to mean the capacity to stimulate a humoral immune response by activating the B lymphocytes so as to amplify the production of antibodies directed against the papillomavirus antigens or to stimulate a cell-mediated immune response by activating the T lymphocytes so as to
20 trigger a significant cytotoxic response against tumor cells or cells infected by a papillomavirus. As a guide, the immunostimulation can be evaluated in an animal model (by comparing the level of rejection in an animal into which a tumor expressing the target antigen has been transplanted, this being in the presence and in the
25 absence of the immunostimulant). More generally, the means for demonstrating an immunostimulation are indicated in Roitt (in *Immunology*, 4th edition, Moby Ltd).

It is possible to use a native immunostimulatory
30 molecule as found in a mammal, and, in particular, in man, a portion thereof, a chimeric molecule obtained from the fusion of sequences of various origins or a mutant, provided, however, the immunostimulatory function is preserved. Among all the molecules which can be
35 envisaged, a polypeptide consisting of or derived from interleukin-2, interleukin-7, interleukin-12 and co-adhesion molecules B7.1 and B7.2 will be preferably used, interleukin-2 and the molecule B7.1 being particularly preferred within the framework of the present invention.

A preferred composition according to the invention comprises:

- 5 (1) a polypeptide from the E6 region, a polypeptide from the E7 region, a polypeptide from the L1 region and a polypeptide from the L2 region of a papillomavirus,
- (2) a polypeptide from the E6 region, a polypeptide from the E7 region of a papillomavirus and a polypeptide derived from interleukin-2,
- 10 (3) a polypeptide from the E6 region, a polypeptide from the E7 region of a papillomavirus and a polypeptide derived from the molecule B7.1,
- (4) a polypeptide from the E6 region, a polypeptide from the E7 region of a papillomavirus, a polypeptide derived from the molecule B7.1 and a polypeptide derived from interleukin-2,
- 15 (5) a polypeptide from the E6 region, a polypeptide from the E7 region, a polypeptide from the L1 region, a polypeptide from the L2 region of a papillomavirus and a polypeptide derived from interleukin-2,
- 20 (6) a polypeptide from the E6 region, a polypeptide from the E7 region, a polypeptide from the L1 region, a polypeptide from the L2 region of a papillomavirus and a polypeptide derived from the molecule B7.1, or
- 25 (7) a polypeptide from the E6 region, a polypeptide from the E7 region, a polypeptide from the L1 region, a polypeptide from the L2 region of a papillomavirus, a polypeptide derived from the molecule B7.1 and a polypeptide derived from interleukin-2.

30 Given the observations recalled above on the incidence of infection by some HPV types in the cases of cancer of the neck of the uterus, a composition according to the invention comprises a polypeptide from a risk-carrying papillomavirus of the HPV-16, HPV-18, HPV-31,
35 HPV-33 and/or HPV-45 type, and in particular from the HPV-16 virus. Of course, in the case where the composition includes several papillomavirus antigens, they may be of common or different origin.

In general, a polypeptide from a papillomavirus

or having an immunostimulatory activity can be produced by the conventional methods of chemical synthesis or alternatively by recombinant DNA techniques (see for example Maniatis et al., 1989, *Laboratory Manual*, Cold Spring Harbor, Laboratory Press, Cold Spring Harbor, NY). More particularly, a method of preparation comprises the act of culturing a cell transformed with a DNA fragment coding for the polypeptide in question so as to generate a producing cell and the act of harvesting said polypeptide from the culture. The producing cell may be of any origin and without limitation, a bacterium, a yeast or alternatively a mammalian cell, insofar as the DNA fragment considered is either integrated into its genome or integrated into an appropriate expression vector capable of replicating. Of course, the DNA fragment is placed under the control of transcriptional and translational signals allowing its expression in the producing cell. Expression vectors and control signals are known to persons skilled in the art.

The present invention also relates to a pharmaceutical composition intended for the treatment or prevention of a papillomavirus infection or tumor, which comprises, as therapeutic agent(s), one or more recombinant vectors into which there are inserted DNA fragments coding for:

- (1) at least one polypeptide from the early region of a papillomavirus and at least one polypeptide from the late region of a papillomavirus,
- (2) at least one polypeptide from the early region of a papillomavirus, at least one polypeptide from the late region of a papillomavirus and at least one polypeptide having an immunostimulatory activity, or
- (3) at least one polypeptide from an early or late region of a papillomavirus and at least one polypeptide having an immunostimulatory activity;

said DNA fragments being placed under the control of the elements necessary for their expression in a host cell or organism.

According to this alternative which is in fact preferred, the therapeutic agent is a vector into which there are inserted the DNA fragments coding for the polypeptides from a papillomavirus or immunostimulants as defined above. This type of composition has the advantage of cheap production and high stability under a variety of environmental conditions. In particular, the preservation conditions are less constraining.

The DNA fragments coding for a polypeptide from a papillomavirus can be obtained by cloning, by PCR (Polymerase Chain Reaction) or by chemical synthesis according to the conventional techniques commonly used starting with positive papillomavirus cells obtained from patients or from collections.

The gene coding for a polypeptide having immunostimulatory activity can also be isolated according to standard techniques from the genome of a cell (genomic type) or from the messenger RNAs of a cell in which it is expressed (complementary DNA type). Moreover, the gene in question can code for (i) a soluble molecule, which is either intracellular or secreted into the external medium, or (ii) a molecule anchored in the membrane and therefore present at the surface of the cells expressing it.

A preferred recombinant vector within the framework of the invention is a viral vector into whose genome there have been inserted the abovementioned DNA fragments so as to allow their transfer and their expression in a host cell or organism. A viral vector which can be used within the framework of the present invention may be derived in particular from a poxvirus, an adenovirus, a retrovirus, a herpesvirus or an adeno-associated virus. Advantageously, it may be a nonintegrative vector having an attenuated virulence. Such vectors, as well as the techniques for preparing them, are known to persons skilled in the art.

In the case where an adenoviral vector is used, there will be preferably used a vector which is non-replicative by virtue of the deletion of regions

essential for replication and, in particular, of the majority of the E1 region so as to avoid its propagation within the host organism or the environment. It goes without saying that other regions of the adenoviral genome can be modified or deleted insofar as the defective essential functions are trans-complemented. A preferred adenoviral vector according to the invention will retain the sequences essential for encapsidation, namely the 5' and 3' ITRs (Inverted Terminal Repeat) and the encapsidation region. The various adenoviral vectors as well as the techniques for preparing them are conventional and are described in Graham and Preveet (1991, in *Methods in Molecular Biology*, vol 7, p 109-128; Ed: E.J. Murey, The Human Press Inc.) and in international application WO 94/28152. If a retrovirus is used, the LTRs (Long Terminal Repeat) and the encapsidation sequences will be retained (see for example Naviaux and Verma, 1992, *Current Opinion in Biotechnology* 3, 540-547).

According to an advantageous embodiment, a recombinant viral vector according to the invention is derived from a poxvirus and, in particular, from an avian poxvirus such as the canarypox virus, a fowlpox virus or a vaccinia virus, the latter being preferred. Among all the vaccinia viruses which can be envisaged within the framework of the present invention, the Copenhagen, Wyeth and modified Ankara (MVA for Modified Vaccinia Ankara Virus) strains are preferably chosen. The general conditions for obtaining a vaccinia virus capable of expressing a heterologous gene are taught in European Patent EP 83 286 and application EP 206 920. As for the MVA virus, it is more particularly described in (Mayr et al., 1975, *Infection* 3, 6-14; Sutter and Moss, 1992, *Proc. Natl. Acad. Sci. USA* 89, 10847-10851).

Of course, in the framework of the present invention, the DNA fragments are placed under the control of the elements necessary for their expression. These include the appropriate elements for regulation of transcription as well as signals for initiation and

termination of translation. The promoter is of particular importance. In general, use will be made of a promoter which is functional in the host organism or cell which it is desired to treat and which is suited to the vector used. In addition, it can be modified so as to contain regulatory sequences, for example an element for activating transcription or sequences responding to certain cellular signals. In this regard, it may be advantageous to use a tissue-specific promoter since the lesions associated with papillomaviruses are located at the level of the genital tracts, or a promoter responding to tumor-specific signals (for example which is activated in the presence of growth factors which are generally overexpressed by tumor cells) so as to limit the expression to tumor cells alone.

Among the promoters which can be envisaged within the framework of the invention, there may be mentioned the SV40 (Simian Virus 40) promoters, the HMG (Hydroxy-Methyl-Glutaryl-coenzyme A) promoters, the TK (Thymidine Kinase) promoters, the CMV (cytomegalovirus) promoters, the RSV (Rous Sarcoma Virus) promoters, the adenovirus MLP (Major Late Promoter) promoters which are suited to adenoviral vectors and the Mo-MLV (Moloney Murine Leukemia Virus) LTR promoters which are more specific to retroviral vectors. When the vector is derived from a poxvirus, the promoter will be preferably the promoter of a gene of the poxvirus used, for example the promoter of the gene coding for the protein 7.5K, H5R, TK or K1L of the vaccinia virus. They are described in the literature and can be cloned from the viral genome by conventional techniques.

Moreover, the elements necessary for the expression may also comprise sequences enhancing the expression or maintenance in the host cell (intron, signal sequence, sequence for termination of transcription, site of initiation of translation, sequences modifying the presentation of the polypeptide to the cells of the host's immune system and the like). However, for a vector derived from a poxvirus, the use of introns will be

avoided.

A composition according to the invention can be obtained either with several recombinant vectors, each expressing a given polypeptide, or with a single vector
5 expressing the DNA fragments corresponding to the chosen polypeptides placed under the control of independent or common elements. According to the latter option, there may be used sequences which make it possible to initiate translation internally (IRES) or fusions of the different
10 genes in phase.

The general conditions for obtaining a recombinant vector in use in the present invention are widely described in the state of the art. As regards a poxviral vector, reference may be made to European Patent
15 EP 83 286 the content of which is incorporated herein by reference. These conditions are applicable to the other viruses acceptable as a vector, which possess a non-essential genomic region into which the expression units may be incorporated. Of course, they may be inserted in
20 the same locus or in a different locus. For example, when the Copenhagen strain of a vaccinia virus is used, the preferred site of insertion is the TK locus and/or the K1L locus. The insertion at the level of the viral TK gene has the effect of inactivating it and thus of
25 facilitating the selection of the recombinants. In the context of the MVA strain of a vaccinia virus, the insertion of the immunostimulatory and papillomavirus genes can be carried out within one of the excisions I to VI and, preferably, the excision zone II or III (Meyer et
30 al., 1991, J. Gen. Virol. 72, 1031-1038; Sutter et al., 1994, Vaccine 12, 1032-1040).

In accordance with the objectives pursued by the present invention, a recombinant vector may, in addition, comprise a unit for expression of a selectable marker
35 gene in order to facilitate the steps for isolating and purifying the recombinant virus. There may be mentioned in particular the Neo gene which confers resistance to the antibiotic G418, the pac gene for resistance to puromycin, the herpes simplex virus type 1 (HSV-1) TK

gene which confers sensitivity to certain nucleoside analogs such as ganciclovir or acyclovir, the bacterial genes *LacZ* coding for β -galactosidase and *gus A* coding for β -glucuronidase. The latter two enzymatic markers
5 make it possible to pick out the recombinant viruses by staining in the presence of the substrates X-Gal (5-bromo-4-chloro-3-indolyl β -D-galactopyranoside) and XglcA (5-bromo-6-chloro-3-indolyl β -D-glucoronide) respectively.

10 A pharmaceutical composition according to the invention for the treatment or prevention of a papilloma-virus infection or tumor comprises one or more recombinant vectors derived from the Copenhagen or MVA strain of a vaccinia virus into which there are inserted:

- 15 (1) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus and a DNA fragment coding for the molecule B7.1,
- 20 (2) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus and a DNA fragment coding for interleukin-2,
- 25 (3) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus, a DNA fragment coding for the molecule B7.1 and a DNA fragment coding for interleukin-2,
- 30 (4) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus, a DNA fragment coding for the L1 protein of a papillomavirus and a DNA fragment coding for the L2 protein of a papilloma-virus,
- 35 (5) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus, a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papilloma-virus and a DNA fragment coding for the molecule B7.1,

5 (6) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus, a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papillomavirus and a DNA fragment coding for interleukin-2, or

10 (7) a DNA fragment coding for the E6 protein of a papillomavirus, a DNA fragment coding for the E7 protein of a papillomavirus, a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papillomavirus, a DNA fragment coding for the molecule B7.1 and a DNA fragment coding for interleukin-2.

15 On the other hand, a pharmaceutical composition according to the invention more particularly intended for immunoprophylaxy comprises one or more recombinant vectors derived from the Copenhagen or MVA strain of a vaccinia virus, into which there are inserted:

20 (1) a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papillomavirus and a DNA fragment coding for the molecule B7.1,

25 (2) a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papillomavirus and a DNA fragment coding for interleukin-2, or

30 (3) a DNA fragment coding for the L1 protein of a papillomavirus, a DNA fragment coding for the L2 protein of a papillomavirus, a DNA fragment coding for interleukin-2 and a DNA fragment coding for the molecule B7.1.

35 A composition according to the invention can be prepared according to methods known in the field of vaccines and the applicable doses may vary within a wide range. They depend in particular on the polypeptides and on the virus used, the pathological condition to be treated, the condition of the patient and other parameters which may be evaluated by the clinician. However,

in general, the dose of virus per kilo will be from 10^4 to 10^{11} , advantageously from 10^6 to 10^{10} and, preferably, from 10^7 to 10^9 plaque forming units (pfu) when the therapeutic agent is a viral vector and from 0.05 to 500 mg, advantageously from 0.5 to 200 mg and, preferably, from 1 to 100 mg when the therapeutic agent is of polypeptide origin.

A composition according to the invention can be administered by any conventional route of administration, in particular by the intravenous, intramuscular, subcutaneous or subepithelial route, or alternatively by scarification. In the case of an accessible tumor, it is also possible to use direct injection into the site or in the vicinity of the tumor or to use topical application. As a vaccine, a composition according to the invention can be administered according to practices which are common in the field, for example as a single dose or a dose repeated once or several times after a certain time interval. On the other hand, in the context of a curative treatment, it can be administered frequently for a period which is sufficient for the treatment to be effective. When the therapeutic agent is a viral vector, this virus is preferably in a live form. In the case of a poxviral vector, the use of an attenuated strain such as the thymidine kinase negative Copenhagen strain or MVA strain will be preferred. Finally, a recombinant viral vector can be attenuated by an appropriate chemical treatment known to persons skilled in the art. However, injecting a killed recombinant vector can also be envisaged.

According to a preferred embodiment, a pharmaceutical composition according to the invention comprises a therapeutically effective quantity of a recombinant vector in combination with a pharmaceutically acceptable carrier. The carrier is chosen so as to allow its administration by injection into humans or into animals. It may also comprise a vehicle, a diluent and/or an adjuvant and may be provided in liquid or freeze-dried form.

The present invention also relates to a pharmaceutical composition according to the invention, as a

medicament for the treatment or prevention of cancer of the neck of the uterus, of a dysplasia of the neck of low grade and of a papillomavirus infection.

5 Finally, the present invention also relates to a method of treating or preventing the abovementioned pathological conditions, according to which a pharmaceutically effective quantity of a mixture of polypeptide or of a recombinant vector in use in the present invention is administered to an individual needing such a
10 treatment.

The present invention is illustrated by reference to the following figures.

Figure 1 is a diagrammatic representation of the vector pTG5021 allowing the transfer, to the TK locus of
15 the Copenhagen vaccinia, of the HPV-16 late genes L1 and L2 placed under the control of the promoter p7.5K, the two cassettes being in opposite orientation relative to each other.

Figure 2 is a diagrammatic representation of the vector pTG5065 allowing the transfer, to the K1L locus of
20 the Copenhagen vaccinia, of the HPV-16 early genes E6 and E7 placed under the control of the promoter pH5R (the two cassettes being in opposite orientation relative to each other), of the human IL-2 (IL2h) gene placed under the control of the promoter p7.5K and of the marker gene LacZ
25 (btaGAL) placed under the control of the promoter pK1L.

Figure 3 diagrammatically illustrates the strategy which can be used to introduce the HPV-16 late genes L1 and L2 into the exclusion zone II of an MVA
30 vaccinia virus, the left and right recombination arms being indicated (BRG2 and BRD2) respectively.

EXAMPLES

The present invention is described more fully with the aid of the following examples, without being
35 limited as a result.

The constructions described below are carried out according to the general genetic engineering and

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5 molecular cloning techniques detailed in Maniatis et al.,
(1989, *supra*) or according to the manufacturer's recom-
mendations when a commercial kit is used. Synthetic
oligonucleotide site-directed mutagenesis *in vitro* is
10 carried out with the aid of the kit distributed by
Amersham. PCR amplification techniques are known to
persons skilled in the art (see for example PCR Protocols
- A guide to methods and applications, 1990, published by
Innis, Gelfand, Sninsky and White, Academic Press Inc.).
15 As regards the repair of restriction sites, the technique
used consists in filling the protruding 5' ends with the
aid of the large fragment of DNA polymerase I of *E. coli*
(Klenow).

15 The cloning steps, the recombinant M13 bacterio-
phages are multiplied on the *E. coli* NM522 strain
(Stratagene) in an agar-based minimum medium (7.5% agar)
or in a liquid rich LBM medium. The recombinant plasmids
carrying the ampicillin resistance gene are replicated in
the *E. coli* strains C600 (Stratagene), BJ5183 (Hanahan,
20 1983, J. Mol. Biol. 166, 557-580) and NM522 on agar or
liquid medium supplemented with 100 µg/ml of antibiotic.
The BJ5183 strain is preferably used when the cloning is
carried out by homologous recombination (Bubeck et al.,
1993, Nucleic Acid Res. 21, 3601-3602).

25 The construction of the recombinant vaccinia
viruses is carried out according to the conventional
technology in the field which is disclosed in the
documents already cited and in Mackett et al., (1982,
Proc. Natl. Acad. Sci. USA 79, 7415-7419) and Mackett et
30 al., (1984, J. Virol. 49, 857-864).

EXAMPLE 1: Construction of the Copenhagen vaccinia virus
VVTG5021&5065 expressing the HPV-16 E6, E7, L1
and L2 genes and the human IL-2 gene

A. Isolation of the HPV16 L1 and L2 genes

35 The fragments coding for the L1 and L2 proteins
are isolated by PCR from the genomic DNA of Caski cells

(ATCC 1550) according to general prior art techniques. The fragment for amplification, carrying the L1 sequences, is subcloned into the vector M13TG130 (Kieny et al., 1983, Gene 26, 91-99) to give the construct M13TG8171. The sequence of the cloned L1 gene reveals several mutations compared with the sequence contained in Genbank (accession K02718): C in place of an A at position 248, C in place of an A at position 253, G in place of an A at position 537, G in place of a C at position 682, G in place of an A at position 874, insertion of a triplet ACT at position 1393, deletion of a triplet GAT at position 1390. The insertion of the PCR fragment, carrying the L2 sequences, into the vector M13TG6131 leads to M13TG9126. 5 point mutations are observed compared with the sequence disclosed in Genbank: C in place of a T at position 378, A in place of a G at position 691, A in place of a G at position 702, G in place of an A at position 990 and C in place of an A at position 1092. As a guide, the vector M13TG6131 is derived from M13TG131 (Kieny et al., 1983, *supra*) by mutation of the internal *Bgl*III site situated outside the multiple cloning sites.

B. Construction of the vector pTG5021 for the transfer of the L1 and L2 genes into the TK locus of the genome of the vaccinia virus

The gene coding for the L1 protein is modified by creating a *Bgl*III site upstream of the initiator ATG. The mutated gene is excized from the preceding vector (M13TG8185) by *Bgl*III-*Sac*I digestion and inserted between the *Bam*HI and *Sac*I sites of pTG186-poly. The resulting construct is called pTG4019. The transfer vector pTG186-poly is described in detail in French Patent 2,583,429. It has 10 restriction sites for the insertion of the gene to be transferred and the promoter p7.5K for controlling its expression.

The gene coding for the L2 protein is isolated from M13TG9126 by *Bgl*III-*Hind*III digestion and then cloned

between the *Bam*HI and *Hind*III sites in 3' of the 7.5K promoter. M13TG9127 is obtained into which a *Sal*I site is introduced upstream of the promoter by localized mutagenesis. The expression unit "p7.5K-L2 gene" is isolated from the mutated vector M13TG9129 and cloned into the *Sac*I site of the transfer vector pTG4019 such that the two cassettes p7.5K-L1 and p7.5K-L2 are in opposite orientation. The construct thus obtained pTG5021 is represented in Figure 1. The expression units are transferred into the genome of the Copenhagen strain of the vaccinia virus by homologous recombination. The recombinant virus designated VVTG5021 is isolated by selection with 5-bromodeoxyuridine (5BUDR).

C. Isolation of the HPV16 E6 and E7 genes

The E6 and E7 genes are isolated from the Caski cell line as described in Examples 2 and 3 of European Patent EP 0,462,187. Two constructs were derived from the clone M13E7/E6 containing the HPV-16 E6 and E7 genes so as to facilitate subsequent cloning steps. The first, designated M13TG8188, results from the introduction by site-directed mutagenesis of *Pst*I and *Bam*HI sites upstream and downstream, respectively, of the E7 gene, and the second, M13TG8189, comprises a *Pst*I site upstream of the E6 gene. The introduction of point mutations upstream of an initiator ATG and downstream of a stop codon are within the capability of persons skilled in the art.

The combination of the HPV-16 E7 protein with the product of the retinoblastoma gene has been demonstrated by various authors (see for example Munger et al., 1989, EMBO J. 8, 4099-4105) and correlated with its transforming power. For obvious safety reasons, a nononcogenic mutant, from which the sequences coding for amino acids 21 to 26 of the native E7 protein which are involved in the transformation function have been deleted, is generated by site-directed mutagenesis of the vector M13TG8188 with the aid of the oligonucleotide oTG5118

(SEQ ID NO: 1). M13TG9104 is obtained. The mutated E7 gene is designated hereinafter E7*.

Likewise, it has been demonstrated that the HPV-16 E6 protein could interact with the product of expression of the tumor suppressor gene *p53* (Crook et al., 1991, Cell 67, 547-556). The domain involved in this interaction has been clearly defined and is situated between residues 111 to 115 of the native protein. The vector M13TG9125 is generated by mutagenesis of M13TG8189 with the aid of the oligonucleotide oTG5377 (SEQ ID NO: 2). The mutated E6 gene is designated hereinafter E6*.

D. Construction of the transfer vector pTG5065 carrying the HPV-16 E6 and E7 genes and the human IL-2 gene which are integrated into the K1L locus.

The 5.2 kb *EcoRI* K fragment of the left end of the Copenhagen vaccinia virus genome (Guillard et al., 1985, J. Virol. 53, 316-318) is cloned into the plasmid pUC8 (Gibco BRL) linearized with *EcoRI*. The vector thus obtained is then subjected to controlled digestion with *BglII* followed by ligation so as to delete an 855 bp fragment coding for the K1L host restriction gene (Guillard et al., 1986, Proc. Natl. Acad. Sci., USA 83, 5573-5577). The 3.4 kb *BglII* fragment is isolated from this intermediate construct designated pUC8Khr and then cloned into the *BamHI* site of the plasmid pUC7 (Gibco BRL). pBAC1 is generated, into which an *XhoI* adaptor is introduced on the left of the unique *BglII* site. After digesting with *XhoI* and *BglII*, a *SalI*-*BglII* fragment carrying the p7.5K vaccinia promoter is inserted. The two *EcoRI* sites are removed by *EcoRI* digestion followed by treatment with Klenow and religation. The vector pTG2147 is obtained by introducing into the unique *BglII* site a multiple cloning site isolated from the vector p poly II (Lathe et al., 1987, Gene 57, 193-201) isolated in the form of a *BglII*-*BamHI* fragment. It therefore comprises the recombination arms allowing insertion into the K1L

locus surrounding the p7.5K promoter followed by the restriction sites.

5 The E6* and E7* genes are cloned downstream of the vaccinia promoter pH5R contained in the vector M13TG9132, to give M13TG9138 and M13TG9139, respectively. As a guide, the vector M13TG9132 comes from the insertion of the promoter of the H5R gene isolated by PCR from the viral genome into the phage M13TG6131.

10 Moreover, the cDNA coding for human interleukin-2 is isolated from the plasmid pTG36 (French Patent 2,583,770), introduced into an intermediate vector and taken out again by *SalI*-*BamHI* digestion so as to be inserted into the transfer vector pTG2147 targeting the locus K1L. The insertion is made at the level of the *PstI* and *XbaI* sites situated downstream of the promoter p7.5K, with the aid of cloning adaptors. pTG5056 is obtained.

15 The expression unit pH5R-E6* is isolated from the vector M13TG9139 in the form of a *BglIII*-*BamHI* fragment which is introduced into the *BamHI* site of the vector pTG5056. The vector pTG5060 is generated. The expression unit pH5R-E7* is purified from M13TG9138 and inserted at the *BamHI* site of the vector pTG5060, to give pTG5062. It is indicated that insertion at the K1L locus leads to recombinant viruses having a replication capacity which is reduced or even destroyed in certain cell types. For these reasons, a positive selectable marker is introduced to facilitate identification of the recombinant viruses. The vector pTG5065 results from the cloning, into the *BamHI* site of pTG5062, of an expression cassette "pK1L-LacZ gene" isolated from the plasmid pIV75 (Chen et al., 1993, Virology 196, 682-693) by *BglIII*-*BamHI* cleavage. As visualized in Figure 2, it allows the transfer of the E6*, E7* and IL-2 units into the K1L locus of the vaccinia virus.

30 The recombinant vaccinia virus, called VVTG5065, is generated by homologous recombination after transfection of the vector pTG5065 into chicken embryo cells infected with the wild-type Copenhagen vaccinia and picked out by staining under agar with X-Gal. The vaccinia

viruses VVTG5021 and VVTG5065 are used in double infection tests. The double recombinants are selected according to the two markers: formation of blue plaques in the presence of X-Gal and deletion of the TK marker. These, called VVTG5021&5065, express the units p7.5K-IL-2, pH5R-E6* and pH5R-E7* integrated at the K1L locus and the units p7.5K-L1 and p7.5K-L2 integrated at the TK locus.

The expression of the HPV genes can be verified by Western-blot analysis with the aid of appropriate monoclonal or polyclonal antibodies or by inverse PCR. The production of IL-2 can be evaluated by ELISA test or by CTL test as described in the literature. As a guide, the level of expression in vitro is of the order of 60 ng/ml/10⁶ cells infected with 1 pfu/cell and 24h.

EXAMPLE 2: Construction of the Copenhagen vaccinia virus expressing the E6 and E7 genes and the human IL-2 gene

The cDNA coding for human interleukin-2 is isolated from the plasmid pTG36 by *Pst*I digestion and inserted into the *Pst*I site of the plasmid pTG186, giving pTG188. The virus obtained by homologous recombination is called VVTG188.

The E6* gene is isolated from M13TG9125 in the form of a *Pst*I-*Bam*HI fragment and inserted downstream of the 7.5K promoter between the *Pst*I and *Xba*I sites of pTG2147 in the presence of a *Bam*HI-*Xba*I adaptor, giving pTG5057. The E7* gene is placed under the control of the H5R vaccinia promoter producing M13TG9138 and the cassette bordered by the *Bam*HI-*Bgl*III sites is subcloned into the *Bam*HI site of pTG5057. pTG5059 is obtained, into whose *Bam*HI site the *LacZ* selectable marker is inserted under the control of the promoter of the K1L vaccinia gene by *Bgl*III-*Bam*HI digestion of the vector pIV75. The resulting construct is designated pTG5061 and the virus generated by homologous recombination with the vaccinia genome VVTG5061.

A recombinant vaccinia virus expressing the E6*

and E7* genes integrated at the K1L locus and the human IL-2 gene integrated at the TK locus is obtained by co-infection of chicken embryo cells with the viruses VVTG5061 and VVTG188. As a guide, the latter produces a quantity of IL-2 approximately greater than 400 ng per 10⁶ cells infected at 1 pfu per cell for 24h. The doubly recombinant virus is designated VVTG5061&188.

EXAMPLE 3: Construction of a Copenhagen vaccinia virus expressing the E6 and E7 genes, and the gene coding for the human adhesion cofactor B7.1

The cDNA coding for B7.1 is isolated from an mRNA preparation obtained from the Daudi cell line (ATCC CCL-213) by inverse PCR using the primers oTG6353 and oTG6352 (SEQ ID NO: 3 and 4). It is indicated that the sequence of the gene is disclosed in Genebank under accession number M27533. The amplified fragment is cloned between the BglII and EcoRI sites of M13TG6131 leading to M13TG9149 and then between the BamHI and EcoRI sites of pTG186. The construct is called pTG5090 and the virus obtained by homologous recombination VVTG5090. A vaccinia virus expressing the HPV-16 E6* and E7* genes integrated at the K1L locus and the human B7.1 gene integrated at the TK locus can be obtained by co-infection of chicken embryos with the viruses VVTG5061 and VVTG5090. The recombinant virus is designated VVTG5061&5090.

EXAMPLE 4: Construction of an MVA strain of a vaccinia virus expressing the E6 and E7 genes and the B7.1 gene integrated into the zone of excision III

The MVA virus is derived from the Ankara strain of the vaccinia virus. It is not capable of generating infectious particles on mammalian cells but develops satisfactorily on chicken embryo fibroblasts. Its adaptation to these cells caused the excision of 6 regions which are not essential for its development and its

infectious cycle on this type of cells (disappearance of about 15% of the viral genome; Meyer et al., 1991, J. Gen. Virol. 72, 1031-1038). Exogenous genetic material can be integrated at the level of any of these excision zones. Within the framework of the present invention, excisions II and III, which are located at the level of the *HindIII* restriction fragments N and A respectively, are used (Altenburger et al., 1989, Arch. Virol. 105, 15-27).

10 In a first instance, the vector pTG6019 allowing insertion into the excision zone III of the MVA virus is constructed. The homologous recombination arms on either side of the excision zone III are isolated by PCR from the viral genome (see American Patent US 5,185,146) and
15 the primers oTG7637 and oTG7638 (SEQ ID NO: 5 and 6) for the left arm and oTG7635 and oTG7636 (SEQ ID NO: 7 and 8) for the right arm. The amplified fragments are cloned into the *EcoRI* site of the vector pTG1E, to give pTG6019. The genetic material to be transferred is inserted
20 between the two recombination arms. The vector pTG1E is similar to pTG1H (French Patent 2,583,429) except for the presence of an *EcoRI* adaptor in place of multiple cloning sites.

Firstly, a cassette for expression of the *gus A* marker gene is inserted. The 7.5K promoter is first
25 cloned into the *BamHI* site of pTG6019. pTG6021 is obtained into whose *BamHI* site the *gus A* gene generated in the form of a *BglIII-BamHI* fragment is inserted. This can be obtained from the sequence disclosed in the
30 literature. The resulting construct is called pTG6022. The presence of the marker will make it possible to discriminate between the wild-type viruses and the recombinant viruses by detecting the GUS enzymatic activity with the substrate XglcA. A red color reveals
35 the β -glucuronidase activity. However, for the purpose of a clinical application, it may be useful to be able to remove this bacterial marker from the final product after selection of the recombinant viruses. To do this, advantage is taken of the capacity of vaccinia to delete

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The units for expression of the HPV-16 late genes can be inserted into the excision zone II with the aid of the transfer vector pTG6018. It is constructed by inserting into the *EcoRI* site of pTG1E left and right recombination arms bordering the II excision, which are generated by PCR using the primers oTG7665 and oTG7584 (SEQ ID NO: 11 and 12) and oTG7639 and oTG7640 (SEQ ID NO: 13 and 14). As above, a cassette for expression of a positive marker is integrated between the two arms in order to facilitate the detection of the recombinant viruses, it being possible to remove it after the selection step (Spehner et al., 1990, J. Virol. 64, 527-533). The vector pTG6020 results from the cloning of the *LacZ* gene placed downstream of the promoter p7.5K into the vector pTG6018 linearized with *BamHI*. pTG6020 is modified by inserting a sequence p7.5K between its *BamHI* and *SacI* sites situated downstream of the *LacZ* gene. pTG6024 is obtained. The cassettes L1 and L2 can then be introduced according to a strategy as shown in Figure 3.

EXAMPLE 5: Construction of an MVA strain of vaccinia virus expressing the E6 and E7 genes, and the IL-2 gene integrated into the excision zone III

The same strategy as above is used to introduce the IL-2 gene into the vector pTG6056. After cloning the recombinant oligonucleotides oTG10503 and oTG10502 (SEQ ID NO: 15 and 16) to produce pTG6076, the latter is cleaved with *XhoI* and the *BglIII-EcoRI* fragment is prepared from M13TG9185 (pH5R-IL-2) inserted by homologous recombination. The vector pTG6090 thus obtained comprises the *gus A* marker gene in a "labile" form, the cassettes for expression of the HPV-16 early genes as well as a human IL-2 gene cassette. The recombinant virus obtained by homologous recombination with the MVA genome is designated MVATG6090. The late genes can be integrated into the excision zone II using pTG6018, as indicated above.

EXAMPLE 6: Validation of the vector VVTG5021&5065 in an animal model

A. *Toxicity studies*

5 Nude mice received 10^7 pfu of VVTG5021&5065 or
10 10^7 pfu of wild-type vaccinia virus by the intravenous,
intramuscular, subcutaneous or intracranial route. Groups
of 5 mice were formed depending on the type of virus
injected and the route of administration and the number
of animals having lesions linked to vaccinia was evaluat-
15 ed 26 days after the injection. The mice which received
the recombinant virus showed no lesion regardless of the
route of administration used, whereas the majority of the
animals treated with the wild-type vaccinia have lesions
whose frequency and seriousness depends on the route of
20 administration. Furthermore, deaths are recorded in the
case of an intravenous and intracranial injection. These
data show that VVTG5021&5065 is attenuated compared with
the wild-type virus and that it does not cause lesions
even after an intracranial injection.

20 **B. *Immunoprophylaxy experiments with VVTG5021&5065***

 C57BL6 mice were vaccinated three times by the
subcutaneous route with 10^7 pfu of VVTG5021&5065. Three
days after the last immunization, these animals are
challenged with 10^3 E7W1 cells implanted subcutaneously.
25 As a guide, the E7W1 cells are obtained from a murine
lymphoma line transfected with a vector expressing the
HPV-16 oncogenic E7 gene. The percentage survival of the
animals as a function of time is compared to that
obtained with control mice treated with 10^7 pfu of a
30 nonrecombinant vaccinia virus VVTG186 (derivative of the
TG186 vector described above). Monitoring of mortality
shows a difference between the two groups. In the control
group, 100% of the animals died at D36 whereas 40% of the
animals vaccinated with VVTG5021&5065 were still alive at
35 D36 and more than 30% at D51.

Thus, the recombinant viruses expressing HPV antigens and an immunostimulatory gene exhibit antitumor activity against tumor cells expressing the HPV-16 E7 oncogene.

5 C. *Immunotherapy experiments*

10 C57BL6 mice are inoculated with 10^3 E7W1 cells implanted subcutaneously (D0). 10^7 pfu of recombinant viruses are then administered, also subcutaneously at D3 and then D6 and finally D9 and the percentage survival of the animals is determined compared with the control animals which received a nonrecombinant virus. Whereas 100% of the control animals died, a notable increase in the survival of the animals injected with a mixture of VVTG5061§5021 and VVTG188 is observed. Similar results
15 are obtained after administration of VVTG5065§5021 and VVTG188.

20 These immunotherapy experiments were repeated using a different tumor model, BMK16myc cells replacing the E7w1 cells. The BMK16myc cells are kidney cells from newborn mice transfected with the HPV-16 genome and the murine c myc gene. The animals are treated with 10^7 MVATG6090 viruses expressing the HPV-16 E6* and E7* genes and the human IL-2 gene. Compared with the controls, the treated mice exhibit a delayed tumor growth up to D15.